



REVIEW

# Biomarkers and schizophrenia: a qualitative review

Joana Ribeiro da Silva<sup>1</sup> and Sara Azevedo Pinto<sup>2,3</sup>

## Abstract

Schizophrenia (SCZ) is a psychiatric disorder with a broad spectrum of biological and clinical manifestations of yet not completely clear pathophysiological mechanisms. Several lines of evidence have been supporting the idea that immunoinflammatory, oxidative, hormonal and cellular dysfunctions are implicated on the biomolecular basis of SCZ. However, accurate diagnosis and selection of appropriate treatments remains challenging as a result of the scarcity of objective tests. Furthermore, there is a compelling need to find biomarkers that could predict drug response and tailor pharmacological treatment, particularly in drug-naïve first episode psychosis (FEP). Hence, numerous technologies have been employed in order to search for SCZ biological markers, but evidence relating them to treatment efficacy is lacking. In this regard, some preliminary data suggest promising results. The current review provides information on: (1) potential biomarkers associated with biological disturbances and (2) biological markers associated with treatment response.

**Keywords:** Schizophrenia, Biomarkers, Treatment.

<sup>1</sup>Medical Student, Faculty of Medicine of Porto University, Porto, Portugal

<sup>2</sup>Department of Physiology and Cardiothoracic surgery, Faculty of Medicine, University of Porto, Porto, Portugal

<sup>3</sup>Psychiatry and Mental Health Clinic, Centro Hospitalar de São João, Porto, Portugal

Citation: Silva et al. Biomarkers and schizophrenia: a qualitative review. International Journal of Clinical Neurosciences and Mental Health 2017; 4:2  
DOI: <https://doi.org/10.21035/ijcnmh.2017.4.2>

Received: 01 Feb 2017; Accepted: 23 Jun 2017; Published: 07 Aug 2017

Correspondence: Joana Maria Ribeiro da Silva  
Praceta Capitão Salgueiro Maia, 34, 4500-117 Espinho, Portugal  
E-mail address: [joanamariasilva@gmail.com](mailto:joanamariasilva@gmail.com)



Open Access Publication Available at <http://ijcnmh.arc-publishing.org>

© 2017 Silva et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.



## Introduction

Schizophrenia (SCZ) is a heterogeneous psychiatric disorder of still unclear pathophysiological mechanisms, resulting from a complex interplay between genetic variants and environmental factors [1, 2]. It affects more than 21 million people worldwide [3], being characterised by heterogeneous clinical symptomatology such as delusions, hallucinations, thought disorders and behavioural and emotional changes, with progressive cognitive decline and deterioration of personality [4].

The current diagnostic criteria for psychotic disorders are based on self-report, behavioural observation and illness course (e.g. first episode vs. multiple episodes, state of remission), lacking biological validation [5-7]. Hence, as there are no reliable medical diagnostic tests available [4, 7], the identification of biological markers for SCZ has become a relevant issue.

Biomarkers are objective, quantifiable characteristics of biological processes, disease states, or responses to treatment [8, 9]. Lack of objective tools to identify common patient characteristics, makes the development of biomarkers critical in psychiatry for the application of a more directed and individualised treatment [8, 10]. A biomarker of therapeutic response will be clinically useful if it is accurate, reproducible, easy to interpret, has an adequate sensitivity and specificity, and is acceptable to the patient [11]. Multiple factors have been assigned potentially responsible for the discrepancy verified among studies, particularly when evaluating biomarkers (for example cytokines) in chronic SCZ patients on long-term antipsychotics. Patients' gender, age, body mass index (BMI), smoking and dietary habits, differences in measurement techniques, in tested material or in the stored period of the sample, sampling at different illness' stages (acute vs. chronic or active phase vs. remission), exposure to different type and dosage of antipsychotics, the length of treatment, the duration of hospitalisation, different clinical presentations, or the biological heterogeneity, were implicated in the aforementioned discrepancies [12, 13]. Accordingly, an important characteristic that should be taken into account when investigating biomarkers in SCZ is the stage of the disease. Research at illness' onset is especially significant, since it avoids the effect of confounding variables [14], such as prior psycho-pharmacological treatment [10]. For example, few studies have investigated the molecular signatures in drug-naïve first episode psychosis (FEP) patients, given the difficulty of collecting biological samples before any antipsychotic drug administration [15]. FEP might be the most critical period of SCZ and possibly the most opportune time for studying key mechanisms that influence treatment response and outcome [10]. In addition, when treated early, FEP patients might have a better response rate to antipsychotics compared to those with longer duration of untreated illness [16, 17].

This qualitative review, by no means exhaustive, based

on online database search on key terms as schizophrenia, biomarkers and treatment, discusses recent and contrasting findings in the fields of immune system and inflammation, oxidative stress (OS), endocrine system, serum metabolomics, genetic and central nervous system (CNS) signatures, which have produced data of interest in identifying biological mediators that may function as potential biomarkers in SCZ.

## Immunoinflammatory biomarkers

The inflammatory response works both as an innate and an adaptive mechanism that allows the organism to deal with a diverse threats. However, under pathological and chronic conditions, the maintenance of this response might become deleterious [14]. Several data support the inflammatory hypothesis in SCZ's pathophysiology. It is characterised by enhanced innate immune response with overproduction and/or imbalance of cytokines, namely an increase in pro-inflammatory and a decrease in anti-inflammatory cytokines [14, 18-20]. Cytokines, key signalling molecules of the immune system, exert their effects by binding specific receptors on a variety of peripheral and brain target cells [21]. Immune changes in the CNS may originate from peripheral immune activation of cytokines that cross the disrupted blood brain barrier (BBB) or that are synthesised by invading immune cells, both of which originate from extraneuronal sources and disturb brain function [18, 22, 23]. Nevertheless, most neuropathologies are most probably due to dysfunction of resident microglia and astrocytes [24].

Although contrasting results can be found, most studies focus on plasma levels of cytokines or T-helper-1 (Th-1) and T-helper-2 (Th-2) imbalance [25-27]. Meta-analyses on serum cytokine differences between acutely relapsed (AR) and drug-naïve FEP patients found significant increases in macrophage-derived cytokines—interleukin (IL)-1 $\beta$ , IL-6, IL-12 and tumor necrosis factor-alpha (TNF- $\alpha$ )—as well as the Th-1-derived cytokines, interferon-gamma (IFN- $\gamma$ ) and IL-2 receptor (sIL-2R) in both groups [21, 28].

The IL-17 signalling pathway was also found to be dysregulated when compared to healthy controls and correlated with symptoms in veterans with SCZ [18]. IL-17 induces the production of factors implicated in the recruitment of monocytes and neutrophils [29]. The levels of very few of those factors were found to be positively or negatively correlated with positive, general and total scores on the Positive and Negative Syndrome Scale (PANSS) in most studies conducted so far [18, 21, 31]. Moreover, a previous study found increased activation of T-helper-17 cells in patients with recent onset SCZ [30]. Therefore, an existing imbalance of the IL-17 pathway, may be implicated in the etiology of SCZ [18, 31].

As changes in cytokine profile come with the onset of the disorder, studying inflammatory biomarkers in

untreated FEP is relevant, since antipsychotic drugs are known to influence their levels [8]. However, few studies have so far assessed the association between cytokine levels and treatment response in FEP [8].

The aforementioned meta-analyses concluded that in AR and FEP patients, increased plasma concentrations of IL-1 $\beta$ , IL-6 and tumor growth factor-beta (TGF- $\beta$ ) normalised with antipsychotic treatment, independently of its direct effects [21]. It was previously found that IL-6 decreased significantly after antipsychotic therapy in FEP and that this change was positively correlated with clinical improvement [32]. Therefore, it was suggested that IL-1 $\beta$ , IL-6 and TGF- $\beta$  might represent disease state markers [21]. In contrast, a more recent longitudinal study demonstrated lack of changes in IL-6 over 12 weeks of antipsychotic treatment in FEP, indicating that it could be a more trait-like marker [33]. As IL-12, IFN- $\gamma$ , TNF- $\alpha$  and sIL-2R increased levels did not normalise with treatment, these were suggested to be disease trait markers [21, 25, 34]. However, a later study did not find any cytokine to function as a trait marker, as all the studied cytokines—IL-6, IL-10, TNF- $\alpha$  and IL-4—initially increased in FEP and normalised after risperidone treatment [35].

A meta-analysis demonstrated in a quantitative manner that a 4 to 8 week-treatment with antipsychotics (typical, atypical or mixed) led to increases in sIL-2R and decreases in IL-1 $\beta$  and IFN- $\gamma$  blood levels [20]. The latter was considered consistent with the anti-inflammatory role associated with antipsychotics [20]. In contrast, in a more recent study, an increase in INF- $\gamma$  levels was observed over 12 weeks of treatment with atypical antipsychotics in FEP patients [33]. Thus, it was suggested that slightly longer treatment induces cytokines' production through the potential of atypical antipsychotics to generate metabolic syndrome [36].

In vitro studies concluded that haloperidol and clozapine decrease cytokine secretion from lymphocytes in both SCZ patients and healthy controls [37]. Increases in the soluble receptors of TNF- $\alpha$ , which attenuate the molecule's pro-inflammatory activity [38], were observed with clozapine, suggesting an anti-inflammatory effect of this agent [39].

A 10-week risperidone treatment in drug-naïve FEP patients had a significant suppressant effect on several serum cytokine levels [35]. A specific cytokine profile indicated that risperidone may normalise monocytic and T-regulatory cell responses [35] and decrease Th-2 functions [31, 35]. The latter contradicts both in vitro studies and the idea of an aggravated inflammatory profile as a consequence of atypical antipsychotics therapy. However, the aforementioned cytokine levels normalisation was not associated with a good clinical response—defined by a 50% reduction in the PANSS total scale—to risperidone [35]. This was suggested to be due to risperidone's specific effects, not secondary to symptomatic improvement [35].

When evaluating the influence of adjunctive therapies in two groups of SCZ patients in remission, the group on

both antipsychotics (typical or atypical) and mood stabilizers showed increased TNF- $\alpha$  levels compared to the group on treated only with antipsychotics (typical or atypical) [40]. However, since in the first group of patients the total PANSS scores reduced by 50%, it was suggested that augmentation with mood stabilizers might exert favourable effects in SCZ [40].

Another field of study includes the cyclooxygenase (COX), an enzyme involved in the fatty acid (FA) metabolism pathway, which produces prostaglandins and mediates inflammation [22]. It was demonstrated that the modulation of COX activity with anti-inflammatory agents such as aspirin [41] and celecoxib [42, 43] as adjunctive therapy to antipsychotics, may improve symptoms in SCZ. 15-Deoxy-Delta-12,14-prostaglandin J<sub>2</sub> (15d-PGJ<sub>2</sub>) is a COX-derived product that exerts its anti-inflammatory effect by targeting peroxisome proliferator-activated receptor-gamma (PPAR- $\gamma$ ) [44]. In FEP patients, 15d-PGJ<sub>2</sub> was found to be decreased in peripheral blood mononuclear cells (PBMCs), compared to healthy controls [14]. Due to its soluble nature 15d-PGJ<sub>2</sub> was set as a potential plasmatic biomarker for FEP [14]. As 15d-PGJ<sub>2</sub>/PPAR- $\gamma$  pathway can be stimulated pharmacologically, it was also suggested as a potential therapeutic target candidate [14].

### Oxidative Stress—a field for new tailored therapies?

The neuroprogression theory of SCZ postulates that changes in the immune system are accompanied by increased inflammatory markers such as tryptophan catabolites and reactive oxygen species (ROS), that affect the growth and function of neuronal circuits since the intra-utero period [45]. Converging evidence indicates that the OS plays a role in SCZ pathophysiology [46-49]. A study on the disturbances in antioxidative defense systems indicated an increase in free radical generation and antioxidant defense impairment in SCZ patients [46]. Therefore, several markers have been employed to assess OS and anti-oxidative status in different diseases, including SCZ [50].

OS converts glucose and lipids to reactive carbonyl compounds, and excess of these are converted into advanced glycation end products (AGEs) and advanced lipoxidation end products [51, 52]. Accumulation of these products is called carbonyl stress, and has been suggested as an environmental factor in the pathophysiology of SCZ [47, 49, 53]. AGEs interact with AGE receptors (RAGE) on the cell membrane, which induces deleterious effects by increasing oxidative and carbonyl stress [49, 54]. A recent cross-sectional and longitudinal study showed altered carbonyl stress markers levels in SCZ [49]. Significantly increased glyceraldehyde-derived AGEs (Glycer-AGEs) levels and decreased soluble form of the RAGE (sRAGE) levels were found in acute SCZ [49]. Although controversial in humans, a counter-regulatory mechanism that abolishes the AGEs-RAGE axis effects was associated with

sRAGE [55]. Therefore, the combination of decreased sRAGE levels with increased Glycer-AGE levels might be a pattern characteristic of SCZ [49]. Glycer-AGEs and Glycer-AGEs/sRAGE ratios were set as significant diagnostic markers for SCZ that distinguished between patients and healthy controls in 70.0% of cases [49].

Sulfhydryl (SH) groups are important antioxidants, responsible for scavenging free radicals and suppress peroxidation [56]. Significant decreases in the levels of total-SH was found in SCZ patients when compared with healthy controls [46], which was explained by possibly increased turnover of these antioxidant defences, in an attempt to prevent oxidative damage in SCZ [46].

A decrease in superoxide dismutase (SOD) enzyme activity, which has antioxidative effects against superoxide anion, was detected in SCZ patients selected from an outpatient clinic [46]. Although this confirms some studies [57, 58], it also contradicts others that found no difference in SOD activity between chronic SCZ patients and healthy controls [59] or found increased SOD activity in FEP and chronic SCZ patients compared to normal controls [60]. As in the latter study, the levels of SOD were greater in FEP than in chronic patients, the authors suggested that OS might occur in the initial phase of SCZ, getting worse with the chronicity of the disorder. This could either be explained by aging or by antipsychotic treatment [60].

Regarding glutathione peroxidase—an OS-inducible enzyme with significant role in the peroxy scavenging mechanism [61]—and catalase—the enzyme that protects the cells from H<sub>2</sub>O<sub>2</sub> accumulation [46]—contrasting results were found. When compared to healthy controls, glutathione peroxidase was found to be decreased and catalase increased in SCZ [46]. Nevertheless, some previous studies reported the opposite [57, 58].

Glutathione reductase activity, essential to maintain the reducing capability of the cell, was found to be significantly increased in SCZ patients compared to normal controls [46], contradicting previous studies [57, 58]. Hence, these results are in line with an existing dysregulation of the oxidant-antioxidant balance in the pathophysiology of SCZ.

In drug-naïve FEP patients, paraoxonase 1 (PON1)—an antioxidant and anti-inflammatory enzyme that protects high-density lipoprotein particles against OS damage [62, 63]—was decreased when compared to healthy controls [48, 62]. Decreased levels of PON1 activity in FEP indicate lowered protection against lipid peroxidation and lowered anti-inflammatory potential [64]. Impaired antioxidant defence and increased levels of the lipid peroxidation product—malondialdehyde—were also detected in the serum of chronic SCZ patients [58, 59]. It was shown that increasing lipid peroxidation accompanies SCZ [65] and further suggested that lowered PON1 activity in the early phase of illness might be associated with increased lipid peroxidation during the course of SCZ [48]. Moreover, the decreased PON1 activity levels in drug-naïve FEP patients was indicated as a key component of the disease

rather than a consequence of antipsychotic treatment [62].

Increased total radical-trapping antioxidant parameter (TRAP)—a marker of total non-enzymatic antioxidant defenses [66]—was also detected in drug-naïve FEP patients [48]. Increased TRAP can possibly be an early compensatory mechanism, which may counterbalance the increase in ROS production [48]. Hence, it was suggested that in an early phase, SCZ may be characterised by lowered PON1 activity and compensatory elevations in non-enzymatic antioxidants [48]. Nonetheless, with disease progression, lipid peroxidation, accumulation of protein damage and decreases in antioxidant levels due to chronic inflammation, lead to OS damage [48, 50].

After treating drug-naïve FEP patients with risperidone, an increase in PON1 activity and a reduction in the marker of lipid peroxidation—lipid hydroperoxide—occurred, indicating an antioxidant effect of risperidone [48]. The latter might be explained by risperidone's specific effects, as those alterations were not significantly associated with clinical response to treatment and nor were they dose-sensitive [48].

A recent meta-analysis reviewed relevant randomised controlled trials that compared the effects of add-on treatment (to standard antipsychotics) of several antioxidants—allopurinol, Ginkgo biloba, N-acetyl cysteine, selegiline, vitamins C and E—with placebo in SCZ [67]. Although most results shown no differences, there was moderate evidence that Ginkgo biloba had a positive effect on psychotic symptoms [67].

The need to evaluate adjunctive antioxidant therapies in SCZ with parallel assessment of oxidative changes before and after treatment is therefore being reinforced.

### Hormones, possible biological markers?

The hypothalamic-pituitary-adrenal (HPA) axis is the main biological system involved in stress response [70]. In FEP patients hyperactivity of the HPA axis was demonstrated [68-70], leading to increased cortisol levels throughout the day, blunted cortisol awakening response (CAR) and decreased cortisol response to psychosocial stressors [71-73]. A reduction in cortisol levels after antipsychotic treatment was associated with an improvement in psychotic symptoms after 12 weeks of follow-up, in both chronic and FEP patients [36, 74]. A longitudinal study conducted in FEP patients showed that those who did not respond to 12-week antipsychotic treatment already had at illness onset a significant lower CAR (measured in the saliva) compared with patients who responded [72]. It was observed that at psychosis onset CAR predicts subsequent treatment response, as blunted CAR shown a tendency to remain unchanged with antipsychotic treatment [75]. On the whole, it is possible that more blunted CAR can possibly be a trait marker, reflecting a more severe illness not modifiable by treatment [33, 72, 75]. It was suggested that future studies should investigate the impact of antiglucocorticoid treat-

ments in psychotic symptoms [33]. Therefore, this would be particularly important in patients with a blunted CAR, thus less likely to respond to antipsychotic treatment [33].

Arginine-Vasopressin (AVP) is classically known for its role as a potent antidiuretic hormone, being also released centrally during stressful experiences [8]. Increased AVP levels were associated with greater positive symptoms and worse verbal learning in female drug-naïve FEP patients, but not in male [76]. In contrast, a further study found AVP levels to be decreased in SCZ patients and their first-degree relatives compared with healthy controls [77]. Therefore, because alterations were observed in patients and relatives, AVP was suggested as a marker of biological vulnerability for SCZ [77].

Apart from its classically known role as a hormone involved in parturition and lactation, oxytocin (OT), also influences behaviours that are typically impaired in SCZ [78]. The risk factors associated with psychotic disorders were related to some extent to a disruption in the ability of physiological levels of OT to modulate social cognition and neuropsychological function [77].

As OT decreases anxiety and neuroendocrine response, while AVP plays an anxiogenic role, interactions between these two hormones, may lead to a shift in positive social behaviour and defensive states observed in SCZ [8].

Accordingly, the above mentioned suggests that it is important to consider neuroendocrine and gender-specific alterations in early FEP.

### Metabolomics—signatures implicated in disease process

Metabolomics refers to the study of metabolism at the global level, involving the complete repertoire of small molecules present in cells, tissues and organisms [79].

Proteomic techniques are becoming increasingly popular for biomarker identification [80]. For example, a recent investigation found higher expression of N-terminal fragments of fibrinogen in serum of first onset, drug-naïve SCZ patients [81]. It was further concluded that these proteins may be useful as biomarkers for SCZ molecular diagnosis.

Cryoglobulins (Cgs) are considered a marker of immune system chronic activation, inflammation, and autoimmune sensitisation [82, 83]. Detectable blood levels of type III (a mixture of polyclonal immunoglobulins) Cgs along with the presence of complement activation split products were found in SCZ patients [84]. The complement cascade hyperactivation in SCZ was demonstrated in a previous study [85] and a complement-dependent mechanism may be implicated in Cgs-induced effects both in vivo and in vitro [86]. An in vitro study shown that type III Cgs, isolated from PBMCs of SCZ patients, may induce the expression of pro-inflammatory and chemotactic cytokines [86]. Thus, type III Cgs contribute to increase blood levels of cytokines in SCZ, being involved in disease-associated peripheral inflammatory responses [19, 86-88].

A number of authors agree that antipsychotics have varying effects on the immune system [89], liver [90] and whole body metabolism [91], which might be manifested by proteomic changes in the circulation. Most proteomic studies attempted to identify changes in protein levels, not considering the effects of post-translational modifications (PTMs). Nevertheless, characteristic changes in PTMs, such as phosphorylation, can be biologically informative [92]. Phosphorylation abnormalities in proteins involved in acute phase response and coagulation pathways were observed in serum from drug-naïve FEP patients compared to healthy controls [93]. More recently, another study was performed in order to investigate the effects of olanzapine on the serum phosphoproteome profile in FEP after 6-weeks treatment [92]. The main effects of olanzapine treatment were changes in the state of protein phosphorylation rather than in protein abundance [92]. Despite no information provided regarding the clinical response, it was suggested that antipsychotic drugs lead to downstream effects, which can be detected in the peripheral circulation, having utility as response biomarkers [92].

Heart-type fatty acid binding protein (H-FABP), is a lipid carrier protein expressed primarily in brain, cardiac and skeletal muscles and the mammary gland [94, 95]. In early stage drug-naïve SCZ patients, it was found that H-FABP serum levels were baseline predictors of response to olanzapine [22]. It was also shown that H-FABP levels and monocyte expression of H-FABP molecular partner—cluster of differentiation 36 (CD36)—were inversely correlated [22]. Patients with low concentrations of H-FABP and high expression of CD36 on monocytes were less likely to respond to olanzapine [22].

MicroRNAs (miRNAs) are 20–25 nucleotide long, single-stranded, nonprotein-coding RNA molecules that regulate protein expression levels [2, 96]. There is evidence of a key role for miRNAs in neurogenesis, neural maturation and brain development [96, 97]. Thus, dysfunction in miRNA signaling contributes to neurodegenerative and psychiatric disorders [2]. A number of circulating miRNAs was quantified in the serum of first-onset SCZ patients and that of healthy individuals [2, 98]. In a recent study aiming to identify miRNAs that are clinically practicable biomarkers, it was observed a significant decrease in miR-132 in the PBMCs of SCZ patients compared with controls [2]. Moreover, miR-132 was previously found to be significantly downregulated in prefrontal cortical tissue in SCZ patients [99]. Taking into account the latter findings, miR-132 might be a superior marker in SCZ. Therefore, serum miRNAs changes can reflect SCZ status and may be used as biomarkers for diagnosing SCZ [98].

Hence, the identified markers might have utility in diagnosing and in critical decision-making regarding antipsychotic treatment.

## Biomarkers and genetics

The most studied risk gene in SCZ is the Disrupted-in-Schizophrenia 1 (DISC1) gene whose function depends on binding to cytoskeleton proteins, such as the Nuclear-distribution gene E homolog like-1 (NDEL1) [100]. Myelin basic protein (MBP) is the only structural myelin protein known to be absolutely required for generating compact myelin sheaths [101]. While longitudinally assessing gene expression in the blood of very early stage drug-naïve FEP patients, NDEL1 and MBP were found to be upregulated [15]. This effect appeared to be attenuated by risperidone treatment [15]. Hence, the authors suggested that both genes NDEL1 and MBP, which are involved in important neuronal processes (neurodevelopment and myelination, respectively), may be putative biomarkers for psychotic disorders [15].

The possibility of cell cycle-related genes to function as biomarkers was tested by measuring and comparing messenger RNA (mRNA) levels in the peripheral blood of SCZ patients and healthy controls [102]. It was found that the combination of cyclin-dependent kinase 4 (CDK4), minichromosome maintenance complex component 7 (MCM7) and DNA polymerase delta 4 accessory subunit (POLD4) mRNA expression might be a potential useful biomarker for SCZ [102]. The latter three genes were significantly decreased in acute SCZ [102]. However, CDK4 expression levels recovered significantly in the remission state [102]. Therefore, it was suggested that the combination of CDK4, MCM7 and POLD4 might be a trait biomarker for SCZ and that CDK4 may be a state biomarker for SCZ [102].

Other genes associated with SCZ are identified in several genome-wide association studies including L-type calcium channel  $\alpha$  subunit type 1 c (CACNA1C), dopamine D2 receptor (DRD2), AMPA receptor subunit 1 (GRIA1), NMDA receptor 2A (GRIN2A), metabotropic glutamate receptor 3 (GRM3), neuregulin 1 (NRG1) and miRNA137 (MIR137), among others [103, 104].

## CNS biomarkers candidates in SCZ

Microglia—the resident macrophages of the brain—are the primary reservoirs of pro-inflammatory cytokines, such as IL-1 $\beta$ , IL-6, TNF- $\alpha$  and INF- $\gamma$  [24, 105]. These macrophages act as antigen presenting cells in the CNS, responding rapidly to even minor pathological changes in the brain [24, 105]. It was hypothesised that subsets of microglia are maintained permanently in an activated or primed state into adulthood as a consequence of a perinatal infection [106]. A subsequent immune challenge in adulthood can cause exaggerated levels of cytokines from primed glia [106]. The microglial hypothesis predicts an increase in pro-inflammatory cytokines by the activated microglia, leading directly to neuronal degeneration, white matter abnormalities and decreased neurogenesis [21, 24].

Thus, this can possibly cause impaired brain function and SCZ symptoms [18, 89]. Recently, it has become possible to quantify microglial activation in vivo using a positron emission tomography ligand that recognizes the translocator protein, a receptor found on activated microglial cells [107, 108]. Binding of one such agent, (R)-[11C]PK11195, was found to be increased, suggesting differential microglial activation, in gray matter [107] and in hippocampus [108] of SCZ patients.

Activated microglia also stimulates astrocytes to produce S100B—an astrocyte-specific cytokine and a marker of inflammation—that is considered to be the equivalent of C-reactive protein in the brain [109, 110]. Increased concentrations of S100B were found in cerebrospinal fluid (CSF) of SCZ patients in an acute psychotic episode compared to matched healthy controls. Serum levels concomitantly measured were also elevated, correlating closely with the CSF concentrations [109]. Another study showed elevated S100B serum levels in both drug-naïve early stage patients and medicated chronic SCZ patients [111], supporting the activation or damage of glial cells in SCZ. Moreover, lower levels of S100B were detected in medicated chronic SCZ patients compared to drug-naïve early stage patients, possibly suggesting that antipsychotic treatment may reduce neurodegeneration or at least the ongoing neuroinflammatory process [111].

The CSF has been a highly informative source of biomarkers in Alzheimer's disease and neuroinflammatory disorders [112]. However, it was less frequently studied in SCZ. CSF biomarkers may be informative by identifying invading immune cells, inflammatory cytokines, pathogens, disease-associated extracellular proteins, alterations in choroid plexus secretory patterns, and diffusion from blood, particularly if the BBB is disrupted [113]. Of the cytokines that are reliably measured in CSF—IL-6, IL-8 and IL-1 $\beta$ —only IL-1 $\beta$  was found to be elevated in the CSF of FEP patients compared to healthy controls [114]. In contrast, in olanzapine-treated chronic SCZ patients, only IL-6 was increased in the CSF compared to normal controls, while IL-1 $\beta$  was undetectable [115]. Moreover, treatment with olanzapine appeared not to influence IL-6, as CSF or serum levels of olanzapine did not correlate with CSF IL-6 [115]. Although unclear, the reason for the discrepancy regarding CSF IL-6 levels in FEP versus chronic SCZ seemed to be related to the chronic progress of the disease [115].

The endogenous cannabinoid receptor agonist, anandamide, reduces inflammation by blocking microglial activation and stimulates neurogenesis [116]. Anandamide concentrations in CSF were found to be 10-fold increased in drug-naïve FEP who were non-heavy cannabis abusers (lifetime users  $\leq$  5 times) when compared to drug-naïve FEP high-frequency users, healthy low-frequency or high-frequency users [116]. Moreover, elevated anandamide concentrations inversely correlated with psychotic symptoms [116]. Elevated concentrations of anandamide were detected exclusively in CSF, making its central origin

more likely [116, 117]. A trend for an association of anandamide elevation in CSF with a longer time to psychosis transition was observed [117]. The latter might indicate that the patients expressing higher CSF anandamide are less likely to have psychotic symptoms [117].

Kynurenic acid (KYNA)—a metabolite of tryptophan and an antagonist of the N-methyl-D-aspartate receptor—is neuroprotective against glutamatergic excitotoxicity but may contribute to psychosis and cognitive impairment [118]. Kynurenine (KYN)—KYNA's precursor—is actively transported into the brain and metabolised by astrocytes to KYNA [118]. In SCZ, elevated levels of KYNA and KYN in the CSF or in the postmortem brain have been consistently reported [119-121]. Because KYNA does not cross the BBB, only CSF levels appear to be informative [113]. It was suggested that increased brain KYNA might constitute a major trigger for cognitive and psychotic symptoms and should encourage the development of biomarkers and novel treatment approaches based on the KYN pathway [115].

A promising new pathway to explore SCZ's molecular mechanisms, seems to be the technology of induced pluripotent stem cells (IPS), derived from peripheral somatic cells of SCZ patients. IPS cells provide a source of human CNS cell lines, including neurons, for discovering biomarkers and testing of target engagement, mechanism and response to medications [122].

## Conclusion

Efforts in identifying and describing biomarkers for SCZ have been claimed by psychiatrists as a way to help inform early diagnosis and monitor the evolution and treatment of the disease. Stratification of SCZ patients could be based on serum molecular profiles [22]. This would allow the adoption of preventive strategies by identifying at-risk individuals, facilitating early intervention, reducing medication non-response, adverse side effects, non-compliance and thereby minimizing morbidity [14, 22]. Furthermore, it would help the development of a personalised medicine approach that is emerging in other areas, such as oncology, increasing the chances of positive therapeutic outcome for each patient [22].

This review identified multiple putative biomarkers and drug targets for SCZ. However, several limitations render it difficult to find reliable biomarkers of the disease and its actual state at each moment. First of all, many of the genetic and immune/inflammatory processes involved in SCZ are reported in other neurological and psychiatric disorders. Second, heterogeneous research methodologies concerning timing, disease phase of the sampled individuals, their gender, age, BMI and other confounders and drug regimens can prevent in-depth conclusions. Most studies do not evaluate clinical responses but only biomarker changes after treatment. Other studies evaluate the statistical association between a specific biomarker and a PANSS subscore only. In addition, different antipsychotics were

used in the same study and between studies. As they have distinct affinities for receptors, they might also have different efficacy biomarkers [8]. Heterogeneous results can be found even when using the same antipsychotic agent, with discrepancies between in vitro and in vivo studies, stages of illness and SCZ different clinical presentations [35]. Moreover, many studies did not take into account patients' baseline symptoms severity. Finally, our conclusions must take into consideration the limitations of this review, particularly the lack of systematic research methods.

Targeting molecular pathways in multi-factorial diseases such as SCZ requires well designed studies, capable of correlating molecular and clinical data, which should be conducted in the future.

## Abbreviations

15d-PGJ2: 15-Deoxy-Delta-12,14-prostaglandin J2; AGE: Advanced glycation end product; AR: Acutely relapsed; AVP: Arginine-Vasopressin; BBB: Blood brain barrier; BMI: Body mass index; CACNA1C: L-type calcium channel  $\alpha$  subunit type 1 c; CAR: Cortisol awakening response; CD36: Cluster of differentiation 36; CDK4: cyclin-dependent kinase 4; Cgs: Cryoglobulins; COX: Cyclooxygenase; CNS: central nervous system; CSF: Cerebrospinal fluid; DISC1: Disrupted-in-Schizophrenia 1; DRD2: Dopamine D2 receptor; FA: Fatty acid; FEP: First episode psychosis; Glycer-AGEs: glyceraldehyde-derived AGEs; GRIA1: AMPA receptor subunit 1; GRIN2A: NMDA receptor 2A; GRM3: metabotropic glutamate receptor 3; H-FABP: Heart-type fatty acid binding protein; HPA: Hypothalamic-pituitary-adrenal; IL: Interleukin; IFN- $\gamma$ : interferon-gamma; IPS: Induced pluripotent stem cells; KYN: Kynurenine; KYNA: Kynurenic acid; MBP: Myelin basic protein; MCM7: Minichromosome maintenance complex component 7; miRNAs: MicroRNAs; mRNA: messenger RNA; NDEL1: Nuclear-distribution gene E homolog like-1; NRG1: neuregulin 1; OS: Oxidative stress; OT: Oxytocin; PANSS: Positive and Negative Syndrome Scale; PBMC: Peripheral blood mononuclear cells; POLD4: DNA polymerase delta 4; PON1: Paraoxonase 1; PPAR- $\gamma$ : Peroxisome proliferator-activated receptor-gamma; PTM: Post-translational modification; RAGE: AGE receptors; ROS: Reactive oxygen species; SCZ: Schizophrenia; sLL-2R: L-2 receptor; SH: Sulfhydryl; SOD: Superoxide dismutase; sRAGE: soluble RAGE; TGF- $\beta$ : Tumor growth factor-beta; Th-1: T-helper-1; Th-2: T-helper-2; TNF- $\alpha$ : Tumor necrosis factor-alpha TRAP: total radical-trapping antioxidant parameter.

## Competing interests

The author declares no conflict of interest.

## References

- Petronis A. The origin of schizophrenia: genetic thesis, epigenetic antithesis, and resolving synthesis. *Biol Psychiatry* 2004; 15;55(10):965-70.
- Yu H, Wu J, Zhang H, Zhang G, Sui J, Tong W, et al. Alterations of miR-132 are novel diagnostic biomarkers in peripheral blood of schizophrenia patients. *Prog Neuropsychopharmacol Biol Psychiatry* 2015; 63:23-9. <https://doi.org/10.1016/j.pnpbp.2015.05.007>
- World Health Organization. Mental Health Schizophrenia. Available from: [http://www.who.int/mental\\_health/management/schizophrenia/en](http://www.who.int/mental_health/management/schizophrenia/en).
- Sadock BJ, Sadock VA. *Synopsis of psychiatry: behavioral sciences/clinical psychiatry*. 10th ed. Philadelphia: Lippincott Williams&Wilkins; 2007.

5. Emsley R, Chiliza B, Asmal L, Harvey BH. The nature of relapse in schizophrenia. *BMC Psychiatry* 2013; 13:50. <https://doi.org/10.1186/1471-244X-13-50>
6. Insel TR, Voon V, Nye JS, Brown BJ, Altevogt BM, Bullmore ET, et al. Innovative solutions to novel drug development in mental health. *Neurosci Biobehav Rev* 2013; 37(10Pt1):2438-44. <https://doi.org/10.1016/j.neubiorev.2013.03.022>
7. van Os J, Kapur S. Schizophrenia. *Lancet* 2009; 374(9690):635-45. [https://doi.org/10.1016/S0140-6736\(09\)60995-8](https://doi.org/10.1016/S0140-6736(09)60995-8)
8. Fond G, d'Albis MA, Jamain S, Tamouza R, Arango C, Fleischhacker WW, et al. The promise of biological markers for treatment response in first-episode psychosis: a systematic review. *Schizophr Bull* 2015; 41(3):559-73. <https://doi.org/10.1093/schbul/sbv002>
9. Strimbu K, Tavel JA. What are biomarkers?. *Curr Opin HIV AIDS* 2010; 5(6):463-6. <https://doi.org/10.1097/COH.0b013e32833ed177>
10. Malhotra AK. Dissecting the heterogeneity of treatment response in first-episode schizophrenia. *Schizophr Bull* 2015; 41(6):1224-6. <https://doi.org/10.1093/schbul/sbv117>
11. Vasan RS. Biomarkers of cardiovascular disease: molecular basis and practical considerations. *Circulation* 2006; 113(19):2335-62. <https://doi.org/10.1161/CIRCULATIONAHA.104.482570>
12. Lv MH, Tan YL, Yan SX, Tian L, Chen DC, Tan SP, et al. Decreased serum TNF-alpha levels in chronic schizophrenia patients on long-term antipsychotics: correlation with psychopathology and cognition. *Psychopharmacology (Berl)* 2015; 232(1):165-72. <https://doi.org/10.1007/s00213-014-3650-y>
13. Zhang XY, Cao LY, Song C, Wu GY, Chen DC, Qi LY, et al. Lower serum cytokines levels in smokers than nonsmokers with chronic schizophrenia on long-term treatment with antipsychotics. *Psychopharmacology (Berl)* 2008; 201(3):283-9. <https://doi.org/10.1007/s00213-008-1295-4>
14. García-Bueno B, Bioque M, Mac-Dowell KS, Barcones MF, Martínez-Cengotitabengoa M, Pina-Camacho L, et al. Pro-/anti-inflammatory dysregulation in patients with first episode psychosis: toward an integrative inflammatory hypothesis of schizophrenia. *Schizophr Bull* 2014; 40(2):376-87. <https://doi.org/10.1093/schbul/sbt001>
15. Ota VK, Noto C, Santoro ML, Spindola ML, Gouvea ES, Carvalho CM, et al. Increased expression of NDEL1 and MBP genes in the peripheral blood of antipsychotic-naïve patients with first-episode psychosis. *Eur Neuropsychopharmacol* 2015; 25(12):2416-25. <https://doi.org/10.1016/j.euroneuro.2015.09.013>
16. Kahn RS, Sommer IE. The neurobiology and treatment of first-episode schizophrenia. *Mol Psychiatry* 2015; 20(1):84-97. <https://doi.org/10.1038/mp.2014.66>
17. Lieberman JA, Tollefson G, Tohen M, Green AI, Gur RE, Kahn R, et al. Comparative efficacy and safety of atypical and conventional antipsychotic drugs in first-episode psychosis: a randomized, double-blind trial of olanzapine versus haloperidol. *Am J Psychiatry* 2003; 160(8):1396-404. <https://doi.org/10.1176/appi.ajp.160.8.1396>
18. Dimitrov DH, Lee S, Yantis J, Valdez C, Paredes RM, Braidia M, et al. Differential correlations between inflammatory cytokines and psychopathology in veterans with schizophrenia: potential role for IL-17 pathway. *Schizophr Res* 2013; 151(1-3):29-35. <https://doi.org/10.1016/j.schres.2013.10.019>
19. Müller N, Schwarz MJ. Immune system and schizophrenia. *Curr Immunol Rev* 2010; 6(3):213-220. <https://doi.org/10.2174/157339510791823673>
20. Tourjman V, Kouassi É, Koué MÈ, Rocchetti M, Fortin-Fournier S, Fusar-Poli P, et al. Antipsychotics' effects on blood levels of cytokines in schizophrenia: a meta-analysis. *Schizophr Res* 2013; 151(1-3):43-7. <https://doi.org/10.1016/j.schres.2013.10.011>
21. Miller BJ, Buckley P, Seabolt W, Mellor A, Kirkpatrick B. Meta-analysis of cytokine alterations in schizophrenia: clinical status and antipsychotic effects. *Biol Psychiatry* 2011; 70(7):663-71. <https://doi.org/10.1016/j.biopsych.2011.04.013>
22. Tomasik J, Schwarz E, Lago SG, Rothermundt M, Leweke FM, van Beveren NJ, et al. Pretreatment levels of fatty acid handling proteins H-FABP and CD36 predict response to olanzapine in recent-onset schizophrenia patients. *Brain Behav Immun* 2016; 52:178-86. <https://doi.org/10.1016/j.bbi.2015.10.019>
23. Watanabe Y, Someya T, Nawa H. Cytokine hypothesis of schizophrenia pathogenesis: evidence from human studies and animal models. *Psychiatry Clin Neurosci* 2010; 64(3):217-30. <https://doi.org/10.1111/j.1440-1819.2010.02094.x>
24. Monji A, Kato T, Kanba S. Cytokines and schizophrenia: microglia hypothesis of schizophrenia. *Psychiatry Clin Neurosci* 2009; 63(3):257-65. <https://doi.org/10.1111/j.1440-1819.2009.01945.x>
25. Kim YK, Myint AM, Verkerk R, Scharpe S, Steinbusch H, Leonard B. Cytokine changes and tryptophan metabolites in medication-naïve and medication-free schizophrenic patients. *Neuropsychobiology* 2009; 59(2):123-9. <https://doi.org/10.1159/000213565>
26. Potvin S, Stip E, Sepehry AA, Gendron A, Bah R, Kouassi E. Inflammatory cytokine alterations in schizophrenia: a systematic quantitative review. *Biol Psychiatry* 2008; 63(8):801-8. <https://doi.org/10.1016/j.biopsych.2007.09.024>
27. Schwarz MJ, Chiang S, Müller N, Ackenheil M. T-helper-1 and T-helper-2 responses in psychiatric disorders. *Brain Behav Immun* 2001; 15(4):340-70. <https://doi.org/10.1006/brbi.2001.0647>
28. Uptegrove R, Manzanares-Tezon N, Barnes NM. Cytokine function in medication-naïve first episode psychosis: a systematic review and meta-analysis. *Schizophr Res* 2014; 155(1-3):101-8. <https://doi.org/10.1016/j.schres.2014.03.005>
29. Aggarwal S, Gurney AL. IL-17: prototype member of an emerging cytokine family. *J Leukoc Biol* 2002; 71(1):1-8.
30. Drexhage RC, Hoogenboezem TA, Cohen D, Versnel MA, Nolen WA, van Beveren NJ, et al. An activated set point of T-cell and monocyte inflammatory networks in recent-onset schizophrenia patients involves both pro- and anti-inflammatory forces. *Int J Neuropsychopharmacol* 2011; 14(6):746-55. <https://doi.org/10.1017/S1461145710001653>
31. Borovcanin M, Jovanovic I, Radosavljevic G, Djukic Dejanovic DS, Bankovic D, Arsenijevic N, et al. Elevated serum level of type-2 cytokine and low IL-17 in first episode psychosis and schizophrenia in relapse. *J Psychiatr Res* 2012; 46(11):1421-6. <https://doi.org/10.1016/j.jpsyres.2012.08.016>
32. Pae CU, Yoon CH, Kim TS, Kim JJ, Park SH, Lee Cu, et al. Antipsychotic treatment may alter T-helper (TH) 2 arm cytokines. *Int Immunopharmacol* 2006; 6(4):666-71. <https://doi.org/10.1016/j.intimp.2005.10.004>
33. Mondelli V, Ciufolini S, Belvederi Murri M, Bonaccorso S, Di Forti M, Giordano A, et al. Cortisol and inflammatory biomarkers predict poor treatment response in first episode psychosis. *Schizophr Bull* 2015; 41(5):1162-70. <https://doi.org/10.1093/schbul/sbv028>
34. Song XQ, Lv LX, Li WQ, Hao YH, Zhao JP. The interaction of nuclear factor-kappa B and cytokines is associated with schizophrenia. *Biol Psychiatry* 2009; 65(6):481-8. <https://doi.org/10.1016/j.biopsych.2008.10.018>
35. Noto C, Ota VK, Gouvea ES, Rizzo LB, Spindola LM, Honda PH, et al. Effects of risperidone on cytokine profile in drug-naïve first-episode psychosis. *Int J Neuropsychopharmacol* 2015; 18(4). <https://doi.org/10.1093/ijnp/pyu042>
36. Zhang XY, Zhou DF, Cao LY, Wu GY, Shen YC. Cortisol and cy-

- tokines in chronic and treatment-resistant patients with schizophrenia: association with psychopathology and response to antipsychotics. *Neuropsychopharmacology* 2005; 30(8):1532-8. <https://doi.org/10.1038/sj.npp.1300756>
37. Song C, Lin Ah, Kenis G, Bosmans E, Maes M. Immunosuppressive effects of clozapine and haloperidol: enhanced production of the interleukin-1 receptor antagonist. *Schizophr Res* 2000; 42(2):157-64. [https://doi.org/10.1016/S0920-9964\(99\)00116-4](https://doi.org/10.1016/S0920-9964(99)00116-4)
  38. Bradley JR. TNF-mediated inflammatory disease. *J Pathol* 2008; 214(2):149-60. <https://doi.org/10.1002/path.2287>
  39. Hinze-Selch D, Deuschle M, Weber B, Heuser I, Pollmächer T. Effect of coadministration of clozapine and fluvoxamine versus clozapine monotherapy on blood cell counts, plasma levels of cytokines and body weight. *Psychopharmacology (Berl)* 2000; 149(2):163-9. <https://doi.org/10.1007/s002139900351>
  40. Dunjic-Kostic B, Jasovic-Gasic M, Ivkovic M, Radonjic NV, Pantovic M, Damjanovic A, et al. Serum levels of interleukin-6 and tumor necrosis factor-alpha in exacerbation and remission phase of schizophrenia. *Psychiatr Danub* 2013; 25(1):55-61.
  41. Laan W, Grobbee DE, Selten JP, Heijnen CJ, Kahn RS, Burger H. Adjuvant aspirin therapy reduces symptoms of schizophrenia spectrum disorders: results from a randomized, double-blind, placebo-controlled trial. *J Clin Psychiatry* 2010; 71(5):520-7. <https://doi.org/10.4088/JCP.09m05117yel>
  42. Akhondzadeh S, Tabatabaee M, Amini H, Ahmadi Abhari SA, Abbasi SH, Behnam B. Celecoxib as adjunctive therapy in schizophrenia: a double-blind, randomized and placebo-controlled trial. *Schizophr Res* 2007; 90(1-3):179-85. <https://doi.org/10.1016/j.schres.2006.11.016>
  43. Müller N, Krause D, Dehning S, Musil R, Schennah-Wolff R, Obermeier M, et al. Celecoxib treatment in an early stage of schizophrenia: results of a randomized, double-blind, placebo-controlled trial of celecoxib augmentation of amisulpride treatment. *Schizophr Res* 2010; 121(1-3):118-24. <https://doi.org/10.1016/j.schres.2010.04.015>
  44. Ray DM, Akbiyik F, Phipps RP. The peroxisome proliferator-activated receptor gamma (PPARgamma) ligands 15-deoxy-Delta12,14-prostaglandin J2 and ciglitazone induce human B lymphocyte and B cell lymphoma apoptosis by PPARgamma-independent mechanisms. *J Immunol* 2006; 177(8):5068-76. <https://doi.org/10.4049/jimmunol.177.8.5068>
  45. Chan RC, Di X, McAlonan GM, Gong QY. Brain anatomical abnormalities in high-risk individuals, first-episode, and chronic schizophrenia: an activation likelihood estimation meta-analysis of illness progression. *Schizophr Bull* 2011; 37(1):177-88. <https://doi.org/10.1093/schbul/sbp073>
  46. Al-Asmari AK, Khan MW. Inflammation and schizophrenia: alterations in cytokine levels and perturbation in antioxidative defense systems. *Hum Exp Toxicol* 2014; 33(2):115-22. <https://doi.org/10.1177/0960327113493305>
  47. Katsuta N, Ohnuma T, Maeshima H, Takebayashi Y, Higa M, Takeida M, et al. Significance of measurements of peripheral carbonyl stress markers in a cross-sectional and longitudinal study in patients with acute-stage schizophrenia. *Schizophr Bull* 2014; 40(6):1366-73. <https://doi.org/10.1093/schbul/sbt234>
  48. Noto C, Ota VK, Gadelha A, Noto MN, Barbosa DS, Bonifácio KL, et al. Oxidative stress in drug naïve first episode psychosis and antioxidant effects of risperidone. *J Psychiatr Res* 2015; 68:210-6. <https://doi.org/10.1016/j.jpsychires.2015.07.003>
  49. Takeda M, Ohnuma T, Takeuchi M, Katsuta N, Maeshima H, Takebayashi Y, et al. Altered serum glyceraldehyde-derived advanced glycation end product (AGE) and soluble AGE receptor levels indicate carbonyl stress in patients with schizophrenia. *Neurosci Lett* 2015; 593:51-5. <https://doi.org/10.1016/j.neulet.2015.03.002>
  50. Zhang M, Zhao Z, He L, Wan C. A meta-analysis of oxidative stress markers in schizophrenia. *Sci China Life Sci* 2010; 53(1):112-24. <https://doi.org/10.1007/s11427-010-0013-8>
  51. Marchbanks RM, Ryan M, Day IN, Owen M, McGuffin P, Whatley SA. A mitochondrial DNA sequence variant associated with schizophrenia and oxidative stress. *Schizophr Res* 2003; 65(1):33-8. [https://doi.org/10.1016/S0920-9964\(03\)00011-2](https://doi.org/10.1016/S0920-9964(03)00011-2)
  52. Prabakaran S, Swatton JE, Ryan MM, Huffaker SJ, Huang JT, Griffin JL, et al. Mitochondrial dysfunction in schizophrenia: evidence for compromised brain metabolism and oxidative stress. *Mol Psychiatry* 2004; 9(7):684-97. <https://doi.org/10.1038/sj.mp.4001532>
  53. Miyashita M, Arai M, Kobori A, Ichikawa T, Toriumi K, Niizato K, et al. Clinical features of schizophrenia with enhanced carbonyl stress. *Schizophr Bull* 2014; 40(5):1040-6. <https://doi.org/10.1093/schbul/sbt129>
  54. Prasad K. Low levels of serum soluble receptors for advanced glycation end products, biomarkers for disease state: myth or reality. *Int J Angiol* 2014; 23(1):11-6. <https://doi.org/10.1055/s-0033-1363423>
  55. Kajikawa M, Nakashima A, Fujimura N, Maruhashi T, Iwamoto A, Matsumoto T, et al. Ratio of serum levels of AGEs to soluble form of RAGE is a predictor of endothelial function. *Diabetes Care* 2015; 38(1):119-25. <https://doi.org/10.2337/dc14-1435>
  56. Vidovic B, Stefanovic A, Milovanovic S, Dordevic B, Kotur-Stevuljevic J, Ivanisevic J, et al. Associations of oxidative stress status parameters with traditional cardiovascular disease risk factors in patients with schizophrenia. *Scand J Clin Lab Invest* 2014; 74(3):184-91. <https://doi.org/10.3109/00365513.2013.873947>
  57. Raffa M, Mechri A, Othman LB, Fendri C, Gaha L, Kerkeni A. Decreased glutathione levels and antioxidant enzyme activities in untreated and treated schizophrenic patients. *Prog Neuropsychopharmacol Biol Psychiatry* 2009; 33(7):1178-83. <https://doi.org/10.1016/j.pnpbp.2009.06.018>
  58. Ranjekar PK, Hinge A, Hedge MV, Ghate M, Kale A, Sitasawad S, et al. Decreased antioxidant enzymes and membrane essential polyunsaturated fatty acids in schizophrenic and bipolar mood disorder patients. *Psychiatry Res* 2003; 121(2):109-22. [https://doi.org/10.1016/S0165-1781\(03\)00220-8](https://doi.org/10.1016/S0165-1781(03)00220-8)
  59. Herken H, Uz E, Ozyurt H, Sogut S, Virit O, Akyol O. Evidence that the activities of erythrocyte free radical scavenging enzymes and the products of lipid peroxidation are increased in different forms of schizophrenia. *Mol Psychiatry* 2001; 6(1):66-73. <https://doi.org/10.1038/sj.mp.400078>
  60. Wu Z, Zhang XY, Wang H, Tang W, Xia Y, Zhang F, et al. Elevated plasma superoxide dismutase in first-episode and drug naïve patients with schizophrenia: inverse associations with positive symptoms. *Prog Neuropsychopharmacol Biol Psychiatry* 2012; 36(1):34-8. <https://doi.org/10.1016/j.pnpbp.2011.08.018>
  61. Chandra R, Aneja R, Rewal C, Konduri R, Dass SK, Agarwal S. An opium alkaloid-papaverine ameliorates ethanol-induced hepatotoxicity: Diminution of oxidative stress. *Indian J Clin Biochem* 2000; 15(2):155-60. <https://doi.org/10.1007/BF02883745>
  62. Brinholi FF, Noto C, Maes M, Bonifácio KL, Brietzke E, Ota VK, et al. Lowered paraoxonase 1 (PON1) activity is associated with increased cytokine levels in drug naïve first episode psychosis. *Schizophr Res* 2015; 166(1-3):225-30. <https://doi.org/10.1016/j.schres.2015.06.009>
  63. Précourt LP, Amre D, Denis MC, Lavoie JC, Delvin E, Seidman E, et al. The three-gene paraoxonase family: physiologic roles, actions and regulation. *Atherosclerosis* 2011; 214(1):20-36. <https://doi.org/10.1016/j.atherosclerosis.2010.08.076>

64. Devarajan A, Shih D, Reddy ST. Inflammation, infection, cancer and all that...the role of paraoxonases. *Adv Exp Med Biol* 2014; 824:33-41.  
[https://doi.org/10.1007/978-3-319-07320-0\\_5](https://doi.org/10.1007/978-3-319-07320-0_5)
65. Dietrich-Muszalska A, Kontek B. Lipid peroxidation in patients with schizophrenia. *Psychiatry Clin Neurosci* 2010; 64(5):469-75.  
<https://doi.org/10.1111/j.1440-1819.2010.02132.x>
66. Pedrini M, Massuda R, Fries GR, de Bittencourt Pasquali MA, Schnorr CE, Moreira JC, et al. Similarities in serum oxidative stress markers and inflammatory cytokines in patients with overt schizophrenia at early and late stages of chronicity. *J Psychiatr Res* 2012; 46(6):819-24.  
<https://doi.org/10.1016/j.jpsychires.2012.03.019>
67. Magalhães PV, Dean O, Andreazza AC, Berk M, Kapczinski F. Antioxidant treatments for schizophrenia. *Cochrane Database Syst Rev*. 2016; 2:CD008919.  
<https://doi.org/10.1002/14651858.cd008919.pub2>
68. Aiello G, Horowitz M, Heggul N, Pariante CM, Mondelli V. Stress abnormalities in individuals at risk for psychosis: a review of studies in subjects with familial risk or with "at risk" mental state. *Psychoneuroendocrinology* 2012; 37(10):1600-13.  
<https://doi.org/10.1016/j.psyneuen.2012.05.003>
69. Belvederi Murri M, Pariante CM, Dazzan P, Heggul N, Papadopoulos AS, Zunszain P, et al. Hypothalamic-pituitary-adrenal axis and clinical symptoms in first-episode psychosis. *Psychoneuroendocrinology* 2012; 37(5):629-44.  
<https://doi.org/10.1016/j.psyneuen.2011.08.013>
70. Borges S, Gayer-Anderson C, Mondelli V. A systematic review of the activity of the hypothalamic-pituitary-adrenal axis in first episode psychosis. *Psychoneuroendocrinology* 2013; 38(5):603-11.  
<https://doi.org/10.1016/j.psyneuen.2012.12.025>
71. Ciufolini S, Dazzan P, Kempton MJ, Pariante C, Mondelli V. HPA axis response to social stress is attenuated in schizophrenia but normal in depression: evidence from a meta-analysis of existing studies. *Neurosci Biobehav Rev* 2014; 47:359-68.  
<https://doi.org/10.1016/j.neubiorev.2014.09.004>
72. Mondelli V, Dazzan P, Heggul N, Di Forti M, Aas M, D'Albenzio A, et al. Abnormal cortisol levels during the day and cortisol awakening response in first-episode psychosis: the role of stress and of antipsychotic treatment. *Schizophr Res* 2010; 116(2-3):234-42.  
<https://doi.org/10.1016/j.schres.2009.08.013>
73. van Venrooij JA, Fluitman SB, Lijmer JG, Kavelaars A, Heijnen CJ, Westenberg HG, et al. Impaired neuroendocrine and immune response to acute stress in medication-naïve patients with a first episode psychosis. *Schizophr Bull* 2012; 38(2):272-9.  
<https://doi.org/10.1093/schbul/sbq062>
74. Garner B, Phassouliotis C, Phillips LJ, Markulev C, Butselaar F, Bendall S, et al. Cortisol and dehydroepiandrosterone-sulphate levels correlate with symptom severity in first-episode psychosis. *J Psychiatr Res* 2011; 45(2):249-55.  
<https://doi.org/10.1016/j.jpsychires.2010.06.008>
75. Bradley AJ, Dinan TG. A systematic review of hypothalamic-pituitary-adrenal axis function in schizophrenia: implications for mortality. *J Psychopharmacol* 2010; 24(4 Suppl):91-118.  
<https://doi.org/10.1177/1359786810385491>
76. Rubin LH, Carter CS, Bishop JR, Pournajafi-Nazarloo H, Harris MS, Hill SK, et al. Peripheral vasopressin but not oxytocin relates to severity of acute psychosis in women with acutely-ill untreated first-episode psychosis. *Schizophr Res* 2013; 146(1-3):138-43.  
<https://doi.org/10.1016/j.schres.2013.01.019>
77. Rubin LH, Carter CS, Bishop JR, Pournajafi-Zazarloo H, Drogos LL, Hill SK, et al. Reduced levels of vasopressin and reduced behavioral modulation of oxytocin in psychotic disorders. *Schizophr Bull* 2014; 40(6):1374-84.  
<https://doi.org/10.1093/schbul/sbu027>
78. Heinrichs M, von Dawans B, Domes G. Oxytocin, vasopressin, and human social behavior. *Front Neuroendocrinol* 2009; 30(4):548-57.  
<https://doi.org/10.1016/j.yfrne.2009.05.005>
79. Smith R, Mathis AD, Ventura D, Prince JT. Proteomics, lipidomics, metabolomics: a mass spectrometry tutorial from a computer scientist's point of view. *BMC Bioinformatics* 2014; 15(Suppl7):S9.  
<https://doi.org/10.1186/1471-2105-15-S7-S9>
80. Lakhan SE. Schizophrenia proteomics: biomarkers on the path to laboratory medicine?. *Diagn Pathol* 2006; 1:11.  
<https://doi.org/10.1186/1746-1596-1-11>
81. Ding YH, Guo JH, Hu QY, Jiang W, Wang KZ. Protein biomarkers in serum of patients with schizophrenia. *Cell Biochem Biophys* 2015; 72(3):799-805.
82. Ramos-Casals M, Stone JH, Cid MC, Bosch X. The cryoglobulinaemias. *Lancet* 2012; 379(9813):348-60.  
[https://doi.org/10.1016/S0140-6736\(11\)60242-0](https://doi.org/10.1016/S0140-6736(11)60242-0)
83. Takada S, Shimizu T, Hadano Y, Matsumoto K, Kataoka Y, Arima Y, et al. Cryoglobulinemia (review). *Mol Med Rep* 2012; 6(1):3-8.  
<https://doi.org/10.3892/mmr.2012.861>
84. Boyajyan A, Khoyetsyan A, Tsakanova G, Sim RB. Cryoglobulins as indicators of upregulated immune response in schizophrenia. *Clin Biochem* 2008; 41(6):355-60.  
<https://doi.org/10.1016/j.clinbiochem.2007.11.014>
85. Mayilyan KR, Weinberger DR, Sim RB. The complement system in schizophrenia. *Drug News Perspect* 2008; 21(4):200-10.  
<https://doi.org/10.1358/dnp.2008.21.4.1213349>
86. Chavushyan A, Hovsepyan M, Boyajyan A. Cryoglobulins as potential triggers of inflammation in schizophrenia. *Schizophr Res Treatment* 2013; 2013:125264.  
<https://doi.org/10.1155/2013/125264>
87. Meyer U, Schwarz MJ, Müller N. Inflammatory processes in schizophrenia: a promising neuroimmunological target for the treatment of negative/cognitive symptoms and beyond. *Pharmacol Ther* 2011; 132(1):96-110.  
<https://doi.org/10.1016/j.pharmthera.2011.06.003>
88. Rothermundt M, Arolt V, Bayer TA. Review of immunological and immunopathological findings in schizophrenia. *Brain Behav Immun* 2001; 15(4):319-39.  
<https://doi.org/10.1006/brbi.2001.0648>
89. Strous RD, Shoenfeld Y. Schizophrenia, autoimmunity and immune system dysregulation: a comprehensive model updated and revisited. *J Autoimmun* 2006; 27(2):71-80.  
<https://doi.org/10.1016/j.jaut.2006.07.006>
90. Choi KH, Higgs BW, Weis S, Song J, Llenos IC, Dulay JR, et al. Effects of typical and atypical antipsychotic drugs on gene expression profiles in the liver of schizophrenia subjects. *BMC Psychiatry* 2009; 9:57.  
<https://doi.org/10.1186/1471-244X-9-57>
91. Correll CU, Lencz T, Malhotra AK. Antipsychotic drugs and obesity. *Trends Mol Med* 2011; 17(2):97-107.  
<https://doi.org/10.1016/j.molmed.2010.10.010>
92. Jaros JA, Rahmoune H, Wesseling H, Leweke FM, Ozcan S, Guest PC, et al. Effects of olanzapine on serum protein phosphorylation patterns in patients with schizophrenia. *Proteomics Clin Appl* 2015; 9(9-10):907-16.  
<https://doi.org/10.1002/prca.201400148>
93. Jaros JA, Martins-de-Souza D, Rahmoune H, Rothermundt M, Leweke FM, Guest PC, et al. Protein phosphorylation patterns in serum from schizophrenia patients and healthy controls. *J Proteomics* 2012; 76 Spec No.:43-55.  
<https://doi.org/10.1016/j.jpro.2012.05.027>
94. Furuhashi M, Hotamisligil GS. Fatty-acid binding proteins: role in metabolic diseases and potential drug targets. *Nat Rev Drug Discov* 2008; 7(6):489-503.  
<https://doi.org/10.1038/nrd2589>

95. Moullé VS, Cansell C, Lugué S, Cruciani-Guglielmacci C. The multiple roles of fatty acid handling proteins in brain. *Front Physiol* 2012; 3:385. <https://doi.org/10.3389/fphys.2012.00385>
96. Jin XF, Wu N, Wang L, Li J. Circulating microRNAs: a novel class of potential biomarkers for diagnosing and prognosing central nervous system diseases. *Cell Mol Neurobiol* 2013; 33(5):601-13. <https://doi.org/10.1007/s10571-013-9940-9>
97. Sun E, Shi Y. MicroRNAs: Small molecules with big roles in neurodevelopment diseases. *Exp Neurol* 2015; 268:46-53. <https://doi.org/10.1016/j.expneurol.2014.08.005>
98. Shi W, Du J, Qi Y, Liang G, Wang T, Li S, et al. Aberrant expression of serum miRNAs in schizophrenia. *J Psychiatr Res* 2012; 46(2):198-204. <https://doi.org/10.1016/j.jpsychires.2011.09.010>
99. Miller BH, Zeier Z, Xi L, Lanz TA, Deng S, Strathmann J, et al. MicroRNA-132 dysregulation in schizophrenia has implications for both neurodevelopment and adult brain function. *Proc Natl Acad Sci U S A*. 2012; 109(8):3125-30. <https://doi.org/10.1073/pnas.1113793109>
100. Hayashi MA, Felicori LF, Fresqui MA, Yonamine CM. Protein-protein and peptide-protein interactions of NudE-Like1 (Ndel1): a protein involved in schizophrenia. *Curr Protein Pept Sci* 2015; 16(8):754-67. <https://doi.org/10.2174/1389203716666150505225251>
101. Weil MT, Möbius W, Winkler A, Ruhwedel T, Wrzoc C, Romanelli E, et al. Loss of myelin basic protein function triggers myelin breakdown in models of demyelinating diseases. *Cell Rep* 2016; 16(2):314-22. <https://doi.org/10.1016/j.celrep.2016.06.008>
102. Okazaki S, Boku S, Otsuka I, Mouri K, Aoyama S, Shirowai K, et al. The cell cycle-related genes as biomarkers for schizophrenia. *Prog Neuropsychopharmacol Biol Psychiatry* 2016; 70:85-91. <https://doi.org/10.1016/j.pnpbp.2016.05.005>
103. Harrison PJ. Recent genetic findings in schizophrenia and their therapeutic relevance. *J Psychopharmacol* 2015; 29(2):85-96. <https://doi.org/10.1177/0269881114553647>
104. Schizophrenia working group of the psychiatric genomic consortium. Biological insights from 108 schizophrenia-associated genetic loci. *Nature* 2014; 511(7510):421-7. <https://doi.org/10.1038/nature13595>
105. Monji A, Kato TA, Mizoquchi Y, Horikawa H, Seki Y, Kasai M, et al. Neuroinflammation in schizophrenia especially focused on the role of microglia. *Prog Neuropsychopharmacol Biol Psychiatry* 2013; 42:115-21. <https://doi.org/10.1016/j.pnpbp.2011.12.002>
106. Bilbo SD, Schwarz JM. Early-life programming of later-life brain and behavior: a critical role for immune system. *Front Behav Neurosci* 2009; 3:14. <https://doi.org/10.3389/neuro.08.014.2009>
107. van Berckel BN, Bossong MG, Boellaard R, Kloet R, Schuitmaker A, Caspers E, et al. Microglia activation in recent-onset schizophrenia: a quantitative (R)-[11C]PK11195 positron emission tomography study. *Biol Psychiatry* 2008; 64(9):820-2. <https://doi.org/10.1016/j.biopsych.2008.04.025>
108. Doorduyn J, de Vries EF, Willemsen AT, de Groot JC, Dierckx RA, Klein HC. Neuroinflammation in schizophrenia-related psychosis: a PET study. *J Nucl Med* 2009; 50(11):1801-7. <https://doi.org/10.2967/jnumed.109.066647>
109. Rothermundt M, Ahn JN, Jörgens S. S100B in schizophrenia: an update. *Gen Physiol Biophys* 2009; 28 Spec No Focus:F76-81.
110. Sen J, Belli A. S100B in neuropathologic states: the CRP of the brain?. *J Neurosci Res* 2007; 85(7):1373-80. <https://doi.org/10.1002/jnr.21211>
111. Zhang XY, Xiu MH, Song C, Chen DC, Wu GY, Haile CN, et al. Increased serum S100B in never-medicated and medicated schizophrenic patients. *J Psychiatr Res* 2010; 44(16):1236-40. <https://doi.org/10.1016/j.jpsychires.2010.04.023>
112. Aluise CD, Sowell RA, Butterfield DA. Peptides and proteins in plasma and cerebrospinal fluid as biomarkers for the prediction, diagnosis, and monitoring of therapeutic efficacy of Alzheimer's disease. *Biochim Biophys Acta* 2008; 1782(10):549-58. <https://doi.org/10.1016/j.bbadis.2008.07.008>
113. Goff DC, Romero K, Paul J, Mercedes Perez-Rodriguez M, Crandall D, Potkin SG. Biomarkers for drug development in early psychosis: current issues and promising directions. *Eur Neuropsychopharmacol* 2016; 26(6):923-37. <https://doi.org/10.1016/j.euroneuro.2016.01.009>
114. Söderlund J, Schröder J, Nordin C, Samuelsson M, Walther-Jallow L, Karlsson H, et al. Activation of brain interleukin-1beta in schizophrenia. *Mol Psychiatry* 2009; 14(12):1069-71. <https://doi.org/10.1038/mp.2009.52>
115. Schwieler L, Larsson MK, Skogh E, Kegel ME, Orhan F, Abdelmoaty S, et al. Increased levels of IL-6 in the cerebrospinal fluid of patients with chronic schizophrenia - significance for activation of the kynurenine pathway. *J Psychiatry Neurosci* 2015; 40(2):126-33.
116. Leweke FM, Giuffrida A, Koethe D, Schreiber D, Nolden BM, Kranaster L, et al. Anandamide levels in cerebrospinal fluid of first-episode schizophrenic patients: impact of cannabis use. *Schizophr Res* 2007; 94(1-3):29-36. <https://doi.org/10.1016/j.schres.2007.04.025>
117. Koethe D, Giuffrida A, Schreiber D, Hellmich M, Schultze-Lutter F, Ruhrmann S, et al. Anandamide elevation in cerebrospinal fluid in initial prodromal states of psychosis. *Br J Psychiatry* 2009; 194(4):371-2. <https://doi.org/10.1192/bjp.bp.108.053843>
118. Javitt DC. Distress intolerance, kynurenic acid, and schizophrenia. *JAMA Psychiatry* 2014; 71(7):749-50. <https://doi.org/10.1001/jamapsychiatry.2014.518>
119. Linderholm KR, Skogh E, Olsson SK, Dahl ML, Holtze M, Engberg G, et al. Increased levels of kynurenine and kynurenic acid in CSF of patients with schizophrenia. *Schizophr Bull* 2012; 38(3):426-32. <https://doi.org/10.1093/schbul/sbq086>
120. Nilsson LK, Linderholm KR, Engberg G, Paulson L, Blennow K, Lindström LH, et al. Elevated levels of kynurenic acid in the cerebrospinal fluid of male patients with schizophrenia. *Schizophr Res* 2005; 80(2-3):315-22. <https://doi.org/10.1016/j.schres.2005.07.013>
121. Sathyaikumar KV, Stachowski EK, Wonodi I, Roberts RC, Rassoulpour A, McMahon RP, et al. Impaired kynurenine pathway metabolism in the prefrontal cortex of individuals with schizophrenia. *Schizophr Bull* 2011; 37(6):1147-56. <https://doi.org/10.1093/schbul/sbq112>
122. Brennand KJ, Simone A, Jou J, Gelboin-Burkhart C, Tran N, Sangar S, et al. Modelling schizophrenia using human induced pluripotent stem cells. *Nature* 2011; 473(7346):221-5. <https://doi.org/10.1038/nature09915>